

Effect of Nasal High Flow Therapy on CO₂ Tension - Physico-mathematical Modelling

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Abstract—The respiratory system of a human embodies complex assembly of tissues and organs (typically internal and external intercostal muscles, diaphragm, lung and rib cage), which are coordinated in a fashion that allows the influx and efflux of air into the airways and lungs. Like all other biological systems, the respiratory system is susceptible to injuries and diseases. Where ventilation has been severely impaired leading to poor gaseous exchange across the lung tissue, biomechanical therapeutic modalities such as continuous positive airway pressure (CPAP) and mechanical ventilators have been prescribed for such patients. Currently, Nasal High Flow therapy (NHFT), a novel ventilation technique has been reported to improve gaseous exchange in both neonates and adults by supplying a constant flow of humidified and warmed air into the lungs. NHFT is presently applied in the management of apnoea of prematurity, respiratory distress syndrome, bronchiolitis, and acute lung injury. In spite of reported success, its mechanisms of action (MOA) are not wholly understood. This work, in terms of relevance, provides some insights into the MOA of NHFT by underscoring the mathematical basis for reported improved gaseous exchange during the administration of NHFT. The mathematical model predictions appreciably agreed with bench-top measurements - indicating 17 % and 24% reduction in end tidal CO₂ concentration upon the respective administration of 30 l/min and 60 l/min NHFT.

Keywords—nasal high flow therapy; capnography; dead space; alveolar CO₂ tension.

I. INTRODUCTION

Capnography is the process of analysing the partial pressure of CO₂ in respiratory gases [1]. Owing to the importance of capnography in current medical practice, medical bodies including the American Association for Respiratory Care [1] and American Society of Anesthesiologists [2] [3] have endorsed it as a method for verifying the correct placement of endotracheal tubes for the provision of respiratory support. Where blood acidosis is clinically diagnosed, capnography may be used to identify the cause by checking for hypercarbia (PaCO₂ > 45 mmHg) [4]. Additionally, since CO₂ is transported from the cells (sites of metabolism) to the lungs via the circulatory system, events such as pulmonary and vascular embolism can be detected using capnography [5].

The CO₂ profile recorded during a breathing cycle is unique in terms of morphology for healthy individuals [6][2]. A CO₂ tension profile for a healthy state is shown in Figure 1. Phase I denotes the baseline where inspiration is about to end. The transition stage, Phase II, physically represents a

blending of alveolar CO₂-rich air and dead space air. Phase III (alveolar plateau) indicates an almost complete saturation of the airway with alveolar air and peaks at point E_tCO₂, known as end-tidal concentration of CO₂. Inspiration begins immediately after E_tCO₂ - marking the commencement of phase IV, where influx of fresh atmospheric air speedily dilutes airway air until the baseline value is reached [6][5][7]. Essentially, deviations from healthy state morphology may be suggestive of a pathological condition of the respiratory system [7][3]. It has been mentioned that changes in baseline level, steadiness of the alveolar plateau and slope of transitional portion may be cardinal to the clinical diagnosis of CO₂ rebreathing and pneumothorax [3][5].

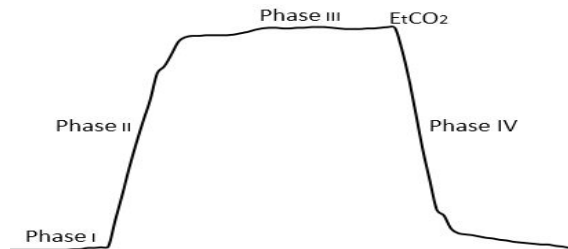


Figure 1. Morphology of a healthy adult capnogram.

Figure 2 shows two polymer models of the upper airway geometry fabricated for experimental work. To fabricate this, a set of computer tomography (CT) images obtained from an adult (age = 44 years and gender = male) was used to reconstruct an *in silico* anatomically representative 3-D model of the upper airway. A detailed description of model making methods has been outlined by Geoghegan et al. [8]. A physical upper airway model, patterned according to the *in silico* model, was built by the use of a 3-D printer (fused deposition modelling type), which utilized acrylonitrile butadiene styrene (ABS) as print material.

Nasal high flow therapy (NHFT) involves the administration of humidified and heated air (up to normothermia) at a constant flow rate. Figure 3 is a pictorial representation of the administration of NHFT via a nasal cannula using Fisher & Paykel Healthcare Airvo2 device. It has been reported that NHFT washes the nasopharyngeal dead space resulting in an increased proportion of inhaled oxygen content and

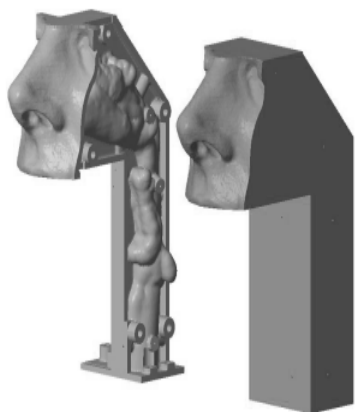


Figure 2. Polymer model of upper airway.

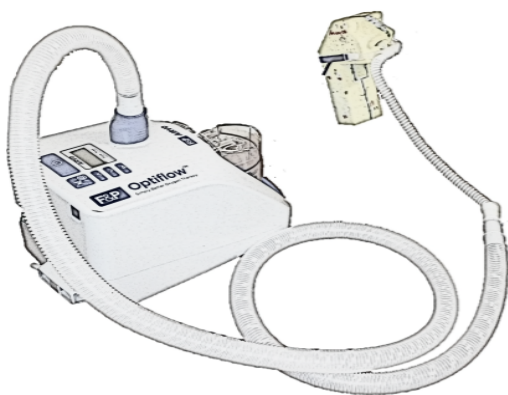


Figure 3. Setup for NHFT administration.

a subsequent improvement in gaseous exchange across the blood-gas barrier [9][10][11]. Arthur et al. [12] pioneered the use of physico-mathematical models to investigate CO₂ fluctuations in the pulmonary system. Their analysis explained the effect of rebreathing on pulmonary CO₂ tension profile. Authors such as Finchman et al. [13] and Milhorn et al [14] used sophisticated compartmental models which included homeostatic response mechanisms, to provide insight into arterial and alveolar reactions to step changes in inhaled CO₂ amounts. Quite recently, Swanson et al. [15] and Benallal et al. [16] applied a two compartment model to shed light on alveolar gas changes during exercise. In the present work, an identical two compartment model has been modified to investigate the amount of CO₂ flushing from the dead space under the influence of NHFT.

In Section 2, formulation of the mathematical model along with the experimental setup is presented. Section 3 comprises of results obtained from both mathematical simulation and bench-top experiment. A discussion of results and concluding observations are presented in Sections 4 and 5 respectively.

II. METHOD

A. Modelling setup

The model presented in this work is identical to that used by Benallal et al. [16] however the inclusion of a constant volumetric flow term to cater for NHFT distinguishes the present model (Figure 4) from it. In this model, the volume of the respiratory system has been thought of as compartmentalized into two main units, namely dead space unit and alveolar unit. The dead space unit represents the volume of all airway regions where gaseous exchange does not occur whilst the alveolar unit denotes the combined volume of lung and airway regions that exchange gas with pulmonary capillaries. Assumptions made during the formulation of model equations include the following (a) administration of NHFT does not change the dead space volume (time invariant) (b) alveolar volume changes due to NHFT is insignificant (c) there is negligible gas loss across the walls of the dead space.

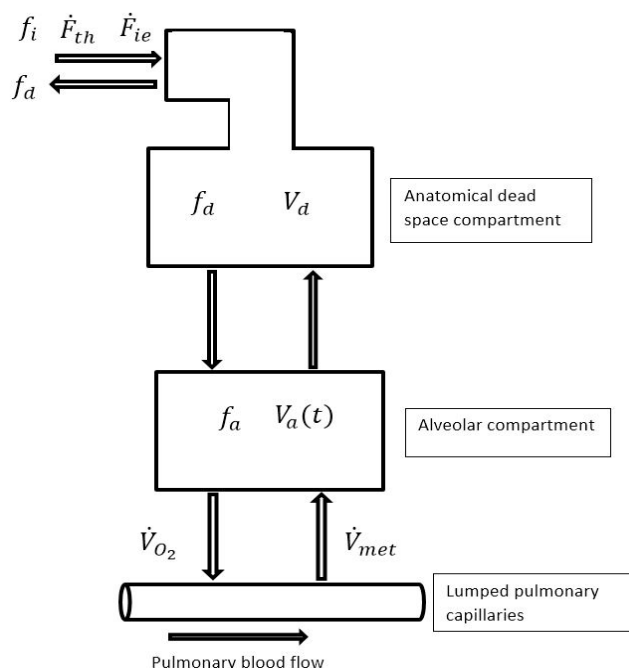


Figure 4. Model representation of respiratory system.

A description of the symbols used in Figure 4 and model equations (1-5) is presented in Table I. Tidal flow data of a healthy male adult (age 23, BMI = 24.6), acquired using a spirometer setup, was used as an input signal to the model. The flow signal was scaled to produce a tidal volume of 500 ml, which is the estimated average for healthy adults [17]. For a resting phase spontaneous breathing of an adult, the functional residual capacity, average metabolic CO₂ production rate (\dot{V}_{met}) and O₂ exchange rate (\dot{V}_{O_2}) across the blood gas barrier were specified as 2500 ml, 240 ml/min and 300 ml/min respectively as used by several authors [16][15][14][5]. Equations (1) and (2) are descriptive of the CO₂ balance in the dead space unit during inspiration and expiration, respectively. In a like manner, the material balance of CO₂ in the alveolar compartment during the breathing cycle is represented by (4) and (5). The alveolar volumetric transience is described by

equation 3. In equations 1, 2, 4 and 5, $\dot{V}_{ie} = \dot{F}_{th} + \dot{F}_i e$.

To solve this 5 non-linear system of equations, a Runge-Kutta (4,5) based solver, ode45, custom-packaged in Matlab (Version 2014b) was employed. The simulation was performed using 50 breathing cycles, which is equivalent to 3.5 minutes of breathing.

TABLE I. MODEL AND EQUATION PARAMETERS.

Symbol	Parameter	Value
f_d	dead space CO ₂ fraction	-
f_a	alveolar CO ₂ fraction	-
\dot{F}_{ie}	ventilatory flow	-
\dot{F}_{th}	NHFT flow	30 and 60 l/min
f_i	inspiratory CO ₂ fraction	-
V_a	alveolar volume (time variant)	-
V_d	dead space volume (constant)	150 ml
\dot{V}_{met}	metabolic CO ₂ influx	240 ml/min
\dot{V}_{O_2}	O ₂ exchange rate	300 ml/min

$$\frac{df_d CO_2}{dt} = \frac{(\dot{V}_{ie})(f_i - f_d)}{V_d} \quad (1)$$

$$\frac{df_a CO_2}{dt} = \frac{\dot{V}_i e (f_a - f_d)}{V_d} \quad (2)$$

$$\frac{dV_a}{dt} = \dot{V}_{ie} + \dot{V}_{met} - \dot{V}_{O_2} \quad (3)$$

$$\frac{df_a CO_2}{dt} = \frac{\dot{V}_{ie} f_d + \dot{V}_{met} - f_a (\dot{V}_{ie} + \dot{V}_{met} - \dot{V}_{O_2})}{V_a} \quad (4)$$

$$\frac{df_a CO_2}{dt} = \frac{\dot{V}_{met} - f_a \dot{V}_{ie} + f_a (\dot{V}_{ie} + \dot{V}_{met} - \dot{V}_{O_2})}{V_a} \quad (5)$$

A sensitivity study was performed to evaluate the independent contribution of the model parameters to $E_t CO_2$. The resolution of the sensitivity scale was limited to a 1 % change in $E_t CO_2$ because a 1 % change significantly affects amount of CO₂ flushed when NHFT is applied. Findings for a 10 % increment on each parameter value is presented in Table II. $E_t CO_2$ was found to be most responsive to initial value of alveolar CO₂ partial pressure (PACO₂) and metabolic CO₂ production rate. Variation in initial dead space CO₂ fraction produced the least change in $E_t CO_2$.

TABLE II. SENSITIVITY STUDY.

Parameter	10% value increment	Change in $E_t CO_2$	Comment
initial PACO ₂	5.86 %	7.71%	very sensitive
\dot{V}_{met}	275 ml	1.06 %	sensitive
initial f_i	0.048 %	0.05 %	less sensitive
FRC	2750 ml	0.12 %	less sensitive
V_d	165 ml	0.43 %	less sensitive
\dot{V}_{O_2}	330 ml/min	0.15%	less sensitive

B. Experimental work

The same physiological flow signal as specified for the mathematical model was programmed into a LabVIEW (Version 8.6) application that operates a pulsatile pump (Figure 5). See component labelling of Figure 5 for setup description. The pulsatile pump (2) connects to a 3-D printed upper airway model via a tubing. NHFT is administered through a nasal cannula (4) by means of a Fisher & Paykel Airvo2 device (3). In performing CO₂ experiments, CO₂ was metered at a bleed rate of 250 ml/min into the pump chamber (piston barrel), allowing a back pressure of 101.3 KPa in the CO₂ source (1). Measurement of CO₂ concentration at the trachea opening of the airway model is performed using a capnograph (5). After a minute, a steady CO₂ profile peaking at $E_t CO_2$ of 5.2% was observed on the computer (6) connected to the capnograph.

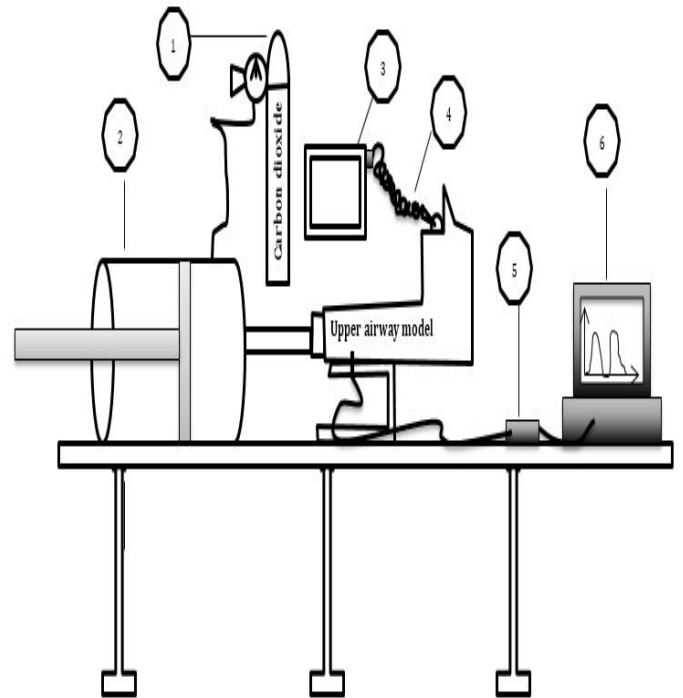


Figure 5. Complete experimental setup

At this point, recording of CO₂ data was performed for 3 minutes followed by 30 l/min NHFT - administered using Fisher & Paykel Healthcare Airvo2 device. Two minutes was allowed for equilibration after which CO₂ recording proceeded for 3 more minutes. An identical procedure was repeated for 60 l/min NHFT, after residual effects from the previous experiment had been eliminated by shutting off CO₂ bleed valve whilst the pump was in operation - allowing CO₂ levels to plummet to near atmospheric values.

III. RESULTS

On Figure 6, plot A shows fractional CO₂ profile in the alveolar unit during a 50 breathing cycle simulation. Transience in breath-by-breath CO₂ fraction is observable up to 60 seconds and then a steady state is reached. The

flow wave used for both simulation and experiment had a period of 4.3 seconds - inspiratory time being 2 seconds and expiratory time being 2.3 seconds. Plot B (Figure 6) represents two cycles of zero-therapy (ZT) ventilatory flow patterns superimposed on corresponding alveolar CO₂ fraction profile at steady state. The distortions on the flow wave at the interface between inspiration and expiration are artefacts introduced by the limited resolution of the spirometer flow device, which is directly linked to the observable local perturbations in fractional alveolar CO₂ profile at the flow transition points. The alveolar CO₂ fraction is seen to rise up to 0.3 seconds into inspiration. Furthermore, over the respiratory cycle, alveolar CO₂ fraction fluctuates between 4.9 % and 5.3 % with a mean value corresponding to 5.1 % .

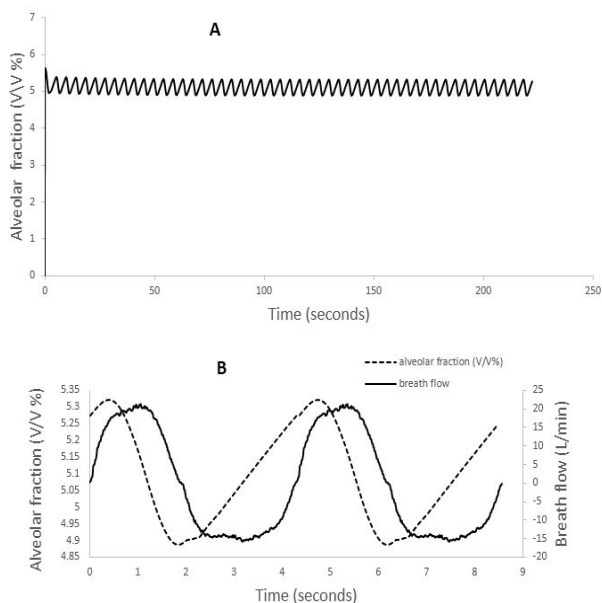


Figure 6. Variation of alveolar CO₂ tension with time.

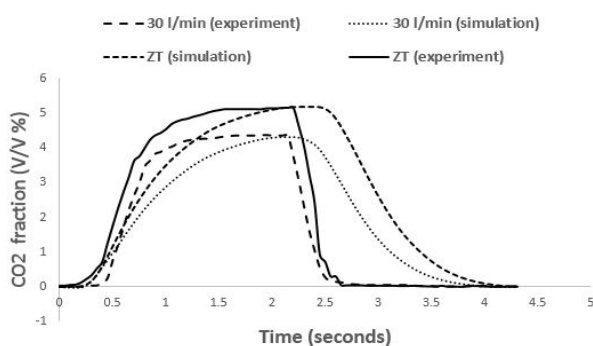


Figure 7. Variation of deadspace CO₂ tension with time.

Figure 7 shows a single respiratory cycle plot of experimental and simulated dead space CO₂ fraction profiles for both ZT and 30 l/min NHFT conditions. When 30 l/min NHFT was applied , dead space E_tCO₂ fell from 5.2 % to about 4.3 % (17% decrement). It is observable from Figure 8 that for 60 l/min NHFT, model simulation under predicts experimental E_tCO₂ by a margin of 8%. In morphology,

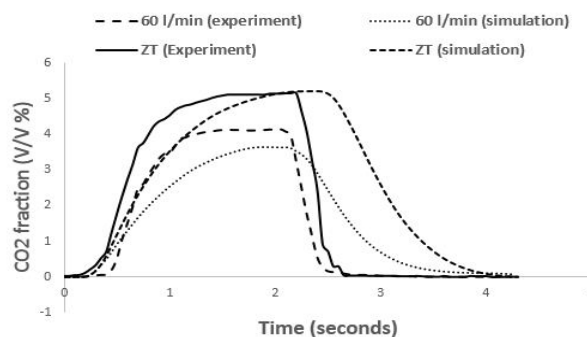


Figure 8. Variation of deadspace CO₂ tension with time.

the simulated dead space CO₂ fraction does not match very well with the experimental measurements. This may be due to the inherent perfect mixing of CO₂ in the dead space compartment (not realistic) which can be made physiological by sub-dividing the dead space volume into several control volumes. The area under the plots (Figures 7 and 8) contain information about dead space CO₂ volume. In quantitative terms, mathematical simulation over-predicts the combined inspiratory and expiratory CO₂ volume for both ZT and 30 l/min NHFT by 8 %. On the same note, CO₂ volume over the respiratory cycle for 60 l/min NHFT is over-predicted by 4%.

IV. DISCUSSION

Generally, the pulsatile pump setup differs from *in vivo* conditions in two aspects, i.e., compliance mismatch (high rigidity of piston barrel) and absence of O₂ exchange mechanism. Since lung elasticity does not enter into the model equations (1-5) and the pulsatile pump is able to deliver the expected tidal volume, the influence of tissue elasticity is eliminated. Given that the ratio of exchanged O₂ volume to tidal volume is quite small (1:25), the present experimental results may therefore considerably approximate the results for the case where an O₂ absorption unit is included in the pulsatile pump assembly. Simulation of no-oxygen condition, however, has indicated a 5 % reduction in expired CO₂ volume.

E_tCO₂ obtained via simulation is comparable to corresponding experimental values for ZT and 30 l/min NHFT. For all cases, slopes of capnogram phases II and IV as predicted by simulation are lesser compared to experimental observations (Figures 7 and 8). This may be due to the model compartmental configuration, which allows for full CO₂ mixing at all times whilst in the experimental setup (same as *in vivo*) there is an established CO₂ front which takes a finite time to travel along the airway. For 60 l/min NHFT, the model under-predicts experimental E_tCO₂ by a change of 8 %. This may be considered as a pronounced effect of the afore-mentioned full CO₂ mixing (not physiological), being heightened by high levels of NHFT.

ZT E_tCO₂ of 5.2 % obtained from simulation corresponds to a tension of 38 mmHg . This value is within physiologic range (37 - 44 mmHg) for healthy adults [18]. The mean alveolar CO₂ fraction of 5.1 % is equivalent to alveolar CO₂ tension (PACO₂) of 39 mmHg, which agrees with the reported physiologic range of 35 mmHg to 45 mmHg [19][20][21]. The rise in alveolar CO₂ fraction for about 0.3 seconds into inspiration is suggestive of rebreathing of CO₂ from the dead

space unit [12].

Over a spontaneous breathing cycle, mean arterial CO_2 pressure (PaCO_2) is approximately the mean of PACO_2 . It has been reported that the lungs can sufficiently engage in oxygen exchange when it is in the state of apnoea, however in this state, CO_2 diffusing across the blood-gas barrier accumulates in the lung and can potentially cause blood acidosis ($\text{PaCO}_2 > 45$ mmHg). Several studies have mentioned an increase in ventilatory rate in severe COPD patients experiencing hypercapnic events ($\text{PACO}_2 > 45$ mmHg), which has been interpreted as a homeostatic reaction to maintain PACO_2 in physiological range [14]. The mean PACO_2 output from the present model may potentially provide some insights into expected PaCO_2 for COPD related hypercapnic events under NHFT conditions. On Figure 7, there is a 17 % fall in $E_t\text{CO}_2$ when 30 liters/min NHFT is applied. The washout volume of CO_2 during expiration, as predicted by the model is 2 ml - representing 15 % of the total expired CO_2 volume for spontaneous breathing. Experimentally, administration of 60 l/min NHFT yielded 24% reduction of ZT $E_t\text{CO}_2$ though the simulated results over-predicts this change. It is however noticeable that the amount of reduction in $E_t\text{CO}_2$ is dependent on flow rate at which NHFT is administered.

Spence et al. [11] used particle imaging velocimetry (PIV) techniques to investigate flow distribution in a silicone upper airway model under NHFT conditions. Their conclusion was that recirculation currents observed in the nasopharynx resulted in CO_2 flushing. Chatila et al. investigated exercise tolerance of severe COPD patients and concluded that NHFT leads to a gain in exercise endurance attributable to increased oxygenation. Flushing of CO_2 may increase the proportion of alveolar ventilation in reference to minute ventilation, thereby boosting oxygen exchange across the blood-gas interface in the lungs [10]. In the light of these reports, the presented two-compartment model, in spite of the outlined limitations, appreciably predicts changes in $E_t\text{CO}_2$ of the bench-top model capnogram for both zero-therapy and NHFT conditions.

V. CONCLUSION

The results from this work show that it is possible to make appreciably satisfactory predictions of the end tidal CO_2 fraction, alveolar CO_2 tension and flushed CO_2 volume for nasal high flow therapy conditions though in terms of morphology, results from the presented two-compartment model show a width-wise disparity from physiologic CO_2 profiles.

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